Review

Beyond blood pressure: new roles for angiotensin II

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Abstract. Since the discovery 100 years ago by Tigerstedt and Bergman of renin, an acid protease generating angiotensin peptide, numerous discoveries have advanced our understanding of the renin-angiotensin system (RAS). The recent cloning of angiotensin receptors and the availability of specific receptor ligands have allowed characterization of angiotensin-receptor-mediated actions, and an increasing number of studies using biochemical, pharmacological and molecular biological methods has focused on the many different physiologi-

cal actions of the RAS in various tissues. Angiotensin II, the main effector peptide of the RAS, exerts most of its known actions in blood pressure control and body fluid homeostasis via the AT_1 receptor. AT_2 receptors not only play a role in growth control and cell differentiation but have been implicated in apoptosis and tissue regeneration. This review focuses on the extrarenal functions of angiotensin, especially in neuronal cells and the nervous system, and on recent advances in angiotensin receptor research.

Key words. Angiotensin II; AT_1 receptors; AT_2 receptors; apoptosis; differentiation; regeneration; signaling pathways.

The renin-angiotensin system

The renin-angiotensin system (RAS) with its effector peptide angiotensin II (ANG II) represents one of the phylogenetically oldest hormone systems and has been a subject of intensive research for more than 100 years [1]. ANG II exerts a variety of actions on different target organs via specific receptors designated AT₁ and AT₂ [2]. The identification of these two main mammalian angiotensin receptors became possible in 1989 with the development of specific receptor ligands for the AT₁ receptor, such as losartan and valsartan, and the AT₂ receptor, such as CGP 42112, PD 123177 and PD 123319 [3].

Most of the known physiological effects of angiotensin, e.g. vasoconstriction, aldosterone release, renal sodium reabsorption as well as central osmoregulatory actions including the release of pituitary hormones into the circulation, have been attributed to the AT₁ receptor. These effects constitute the role of angiotensin peptides as neuromodulator/neurotransmitters in the brain. The AT₁ receptor has, moreover, been shown to mediate cell growth and/or proliferation in various cell types including vascular smooth muscle cells [4, 5], cardiomyocytes [6] and coronary endothelial cells [7]. The contribution of the AT₁ receptor to blood pressure regulation has been examined using AT₁ receptor knock-out mice [8] resulting in a reduction in systolic blood pressure.

The discovery that ANG II was not only a growth promoter (through AT₁ receptors) but also a growth

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inhibitor (mediated by AT_2 receptors) led to a deeper understanding of the RAS in cellular functions, e.g. cell differentiation, tissue regeneration and programmed cell death.

The enzymatic cascade of the RAS starts with the α_2 globulin angiotensinogen which is predominantly synthesized in the liver and constitutively released into the circulation. The aspartyl protease renin, mainly derived from the juxtaglomerular cells in the kidney, generates the biologically inactive angiotensin I (ANG I) in plasma and tissues which is finally converted to ANG II with the help of angiotensin-converting enzyme (ACE), cathepsin D or heart chymase (fig. 1). The octapeptide hormone ANG II exerts its actions via two different receptor subtypes which are characterized in detail below. Finally, activation of several aminopeptidases leads to degradation of angiotensin and to the generation of biologically active metabolites such as ANG III [2-8], ANG IV [3-8] and ANG 1-7 (fig. 1) and subsequently to inactive metabolites.

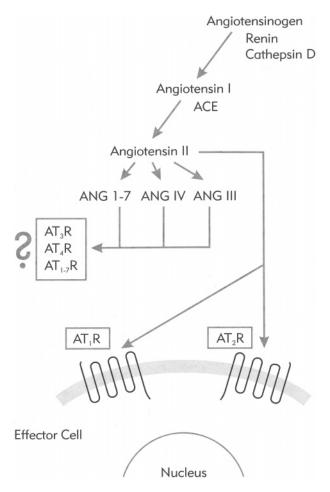


Figure 1. The renin-angiotensin system: pathways for the formation of the angiotensin peptides and their interaction with different angiotensin receptor subtypes.

Angiotensin receptors

Using peptidic and non-peptidic receptor antagonists, different cell surface angiotensin receptor subtypes have been identified, named AT_1 and AT_2 . AT_1 receptors exhibit a high affinity for sartans, e.g. losartan, candesartan, telmisartan, eprosartan and valsartan ($K_i = 10-50$ nM) and a low affinity for CGP 42112A ($K_i > 0.5$ M) and PD 123177 ($K_i > 10$ M). On the other hand, binding sites displaying a high affinity for CGP 42112A ($K_i < 1$ nM) and PD 123177 ($K_i = 10-100$ nM) but a low affinity for sartans ($K_i > 1$ mM) are designated as AT_2 receptors [9]. Several groups have reported that higher concentrations of CGP 42112A exert agonistic effects, thus representing an AT_2 receptor agonist with a high affinity but low potency [7, 10].

Molecular cloning and binding studies have clearly identified two AT_1 receptor isoforms, designated AT_{1a} and AT_{1b} [11, 12]. These isoforms differ in tissue distribution and regulation [13]. Whereas the AT_{1a} receptor seems to be identical with the classical AT_1 receptor, the AT_{1b} receptor displays differences in terms of binding characteristics of selective receptor ligands: PD 123319 > losartan > CGP 42112A [14].

Biochemical data obtained from Siemens et al. [15] also indicate the existence of an additional AT_2 receptor. According to these authors, two different AT_2 receptor isoforms can be discriminated in the neuroblastoma cell line N1E-115 with the help of PD 123319. However, the molecular data point to the existence of a single AT_2 receptor, since its entire coding sequence is localized in one exon without interruption by an intron.

Atypical angiotensin receptors

The development of highly selective angiotensin receptor antagonists also led to the identification of binding sites which differ from those of AT₁ and AT₂ receptors. One example for such a binding site is the so-called AT₄ receptor which exhibits a high affinity for ANG IV (ANG 3–8; cf. fig. 1). Neither losartan nor CGP 42112A and PD 123177 are able to compete for this binding site when reasonable pharmacological concentrations are applied [16, 17].

In endothelial cells, ANG IV stimulates the expression of plasminogen activator inhibitors. Moreover, in cardiac fibroblasts of rabbits, the synthesis of nucleic acids is increased by ANG IV [18]. The AT₄ receptor is expressed in the brain of rabbits and rats where a stimulation of fos immunoreactivity has been detected after ANG IV treatment. This effect could not be suppressed by costimulation with the AT₂ receptor antagonist, PD 123177 [19].

Another atypical binding site for angiotensin has also been described in cell cultures containing 95% human

cardiac fibroblasts [20]. Angiotensin (1-7) > ANG II > ANG IV competitively bound to this site, while losartan and PD 123319 were not able to compete with these agonists. In these cells, angiotensin induced cell proliferation which was not abolished by pretreatment with either losartan or PD 123319. Finally, intracellular receptors have been described which are able to mediate angiotensin effects [21].

Tissue distribution of angiotensin receptors

The angiotensin receptor subtypes have been detected in various tissues [7, 22–30]. In rats, AT_{1a} and AT_{1b} receptors are expressed in the liver and the adrenal glands to almost the same extent. While the AT_{1a} receptor is dominant in vascular smooth muscle cells in the ovary, heart and hypothalamus, the AT_{1b} receptor dominates in tissues involved in central osmoregulation.

In the adult organism, both AT₁ and AT₂ receptor subtypes are expressed in a similar manner in the adrenal glands and the heart [31-45]. On the other hand, the AT₂ receptor is expressed to a greater extent in the uterus, in ovarian granulosa cells, and in distinct areas of the brain, but less in the vascular endothelium [7]. Moreover, the AT₂ receptor is predominantly expressed in fetal tissues, pointing to a role for this receptor subtype in developmental and differentiation processes [46, 47], a function entirely different from the known cardiovascular or volume control of ANG II mediated through the AT₁ receptor. After birth, however, the ratio of AT₁ to AT₂ receptor expression is reversed with the AT₁ receptor subtype dominating in the adult organism; thus, the AT₂ receptor appears to be suppressed in a number of tissues in adult life [for a review see ref. 2]. In the case of pathological incidents like skin lesions [48], post-infarct remodeling [49] or sciatic [50] or optic nerve transection [51], AT₂ receptor expression increases. This receptor has not only been associated with the control of cell growth [2] and—in nervous tissue—with differentiation [52, 53] as well as regeneration [50, 51] and wound repair [54], but has been, furthermore, implicated in glutamate-mediated neuronal cell injury [55, 56] and apoptosis [57, 58]. However, the physiological role of the AT₂ receptor is not yet completely understood and is still the subject of intensive research.

AT₁-receptor-mediated effects

Experimental studies in the last few years have strongly suggested that the RAS plays an important role in the development and maintenance of arterial hypertension. Via its AT₁ receptor, angiotensin not only exerts short-term actions to raise blood pressure, but, furthermore,

evokes by its direct growth-stimulating actions long-term effects leading to renal and cardiovascular pathology, e.g. nephrosclerosis and left ventricular hypertrophy (LVH) and vascular media hypertrophy [for a review see ref. 2]. Therefore, antihypertensive therapy has to focus not only on the minimization of secondary events such as stroke, heart failure or renal disease but also on the avoidance and reduction of structural changes in end organs.

Inhibition of the RAS can be carried out at three different levels (cf. fig. 1):

- 1) Inhibition of renin: although renin acts very specifically on angiotensinogen to generate angiotensin peptides, renin inhibitors have not yet been developed for clinical use due to limited therapeutical potential.
- 2) Inhibition of ACE: apart from generating angiotensin, this enzyme participates in the metabolism of several other peptides, e.g. bradykinin.
- 3) Blockade of AT_1 receptors: the latest most specific approach to oppose angiotensin-mediated actions is the specific antagonization of AT_1 receptors, discussed in more detail below.

All components of the RAS are present in the brain, but they do not coincide in some regions; for example, renin is only reported in low amounts in the cerebral cortex and the basal ganglia, whereas angiotensin and angiotensinogen mRNA can be found in the basal ganglia, septum, amygdala, cortex, hypothalamus and other brain regions [59–62]. Due to the blood-brain barrier, only ANG II endogenously produced in the brain has access to brain ANG II receptors. Neurons express detectable amounts of AT₁ (e.g. hypothalamus, brain stem) and AT₂ receptors (e.g. thalamus, cerebellum, inferior olivary nucleus, retina [28, 51]), but in the circumventricular organs, which have no blood-brain barrier, the high expression of AT₁ receptors seems to be located mostly on glial cells [63].

AT₁ receptor antagonists

The first specific non-peptide AT_1 receptor antagonist clinically introduced as an antihypertensive agent in 1995 was losartan. Additional compounds of this class of drugs followed or will follow in the near future, such as valsartan, irbesartan, eprosartan, candesartan, telmisartan or tasosartan. These compounds are in most cases non-competitive receptor antagonists and do not exert intrinsic activities. They are orally active and exhibit a high selectivity for the AT_1 receptor in various tissues.

For reducing blood pressure, the efficacy of AT_1 receptor antagonists is comparable to that of first-line drugs such as thiazide diuretics, β -blockers, calcium channel blockers and α receptor antagonists. Patients treated

with AT₁ receptor antagonists show a responder rate of 50% or more on monotherapy, which can be further increased by combination with an additional compound of another class, especially diuretics.

The AT₁ receptor antagonists have, moreover, been shown to be well tolerated and, surprisingly, have not so far revealed any class-specific side-effects when used appropriately. Indeed, the number of patients showing unwanted effects or withdrawing from therapy in clinical trials was comparable to patients treated with placebo [64].

An important feature of this substance group is their organ-protective action resulting in a wide range of possible therapeutic advantages in the treatment of hypertension. A few examples are given below.

A tissue-protective action of AT₁ receptor blockade has e.g. been shown in salt-loaded stroke-prone spontaneously hypertensive rats (SHRSP), an animal model of malignant hypertension. In these rats, chronic AT₁ receptor blockade with losartan prevented stroke, malignant nephrosclerosis and cardiac infarction, and increased survival [65–67]. Long-term antihypertensive treatment of SHRSP with losartan can, furthermore, engender beneficial actions on cardiac function and metabolism [68]. Candesartan has been reported to reduce or prevent the development of intima lesions following vascular injury through inhibition of direct and indirect growth-promoting effects of angiotensin in vascular smooth muscle cells [69]. The production of peroxide in human macrophages—which plays a role in the advance of atherosclerosis—can also be decreased by AT₁ receptor blockade, pointing to a beneficial role of AT₁ receptor antagonists in tissue protection [70].

AT₂-receptor-mediated effects in cell differentiation and antiproliferation

In tissue development or remodeling, excessive growth induced by growth factors needs to be controlled by an antiproliferative cellular programme. The two angiotensin receptor subtypes, AT₁ and AT₂, mediate antagonizing effects in terms of growth modulation. AT₂ receptor stimulation inhibits the AT₁-mediated proliferation in various cell types including rat coronary endothelial cells and PC12W cells [7, 52]. Opposing effects of AT₁ and AT₂ receptors on microvascular growth and neointima formation have also been reported [71, 72], and studies from knock-out mice models [73, 74] demonstrated an involvement of AT₂ receptors in blood pressure regulation and behaviour. In adrenal adenomas, a reduction in AT2 receptor mRNA was observed, whereas no consistent differences in AT₁ receptor mRNA were seen [75]. These results suggest a correlation of AT₂ receptors with adrenal tumorgenesis.

Indirect evidence for a role of AT₂ receptors in differentiation and development has been obtained from studies determining the distribution of angiotensin receptor subtypes. During ontogenesis, there is abundant and transient expression of AT₂ receptors in several tissues including the brain [22–24, 28, 76]. In adult animals, the embryonic pattern of high AT₂ and low AT₁ receptor levels is reversed [47, 77]. Due to the transitory abundance of AT₂ receptors in the nervous system, their presence in neuronal cell lines [78, 79], their interaction with T-type calcium currents in non-differentiated NG108-15 cells [80, 81]—calcium playing a crucial role in neuronal differentiation [82]—the involvement of AT₂ receptors in neuronal development has been proposed.

Angiotensin receptor signaling pathways

Analysis of the amino acid sequences of the AT₁ and AT2 receptors has revealed that they belong to the G-protein-coupled 7-transmembrane receptor family. However, the intracellular signaling pathways after receptor stimulation are quite different. The signaling pathways of the AT₁ receptor are now largely understood [83]. Binding of ANG II to the AT₁ receptor leads to the 'classical' G-protein-related cascades with phospholipase (PL)C, PLD and PLA2 stimulation. PLC activation leads to the hydrolysis of phosphatydilinositol 4,5 bisphosphate (PIP2) to inositol triphosphate (IP₃) and diacylglycerol. IP₃ invokes a subsequent increase in intracellular calcium, causing protein kinase C (PKC) activity to increase [84, 85]. The cascade induced by PLD and PLA, leads to the formation of fatty acids, e.g. arachidonic acid, which is a precursor of leukotrienes and prostaglandins. Further research has discovered the contribution of additional signaling mechanisms involving ras/raf-mediated activation of mitogen-activated protein kinases (MAP kinases), the Janus kinase (JAK)/signal transducers and activators of transription (STAT) system, and c-Jun N-terminal kinase family (JNK) activation via tyrosine and serine/ threonine phosphorylation [86]. A neuromodulatory action of angiotensin via stimulation of the AT₁ receptor of the brain norepinephrine system has been reported by Gelband et al. [87]: neuronal synthesis and reuptake of norepinephrine is enhanced via induction of Fos and Jun proteins. In addition to the ras/raf/MAP kinase pathway, the authors recently demonstrated involvement of the protein PKC β subtype and phosphorylation and redistribution of myristolated alanine-rich C kinase substrate (MARCKS) in neurites, leading to persistent stimulation of the neuromodulatory actions of ANG II [88]. Most of these signaling cascades transform the AT₁-receptor-mediated signal into activation

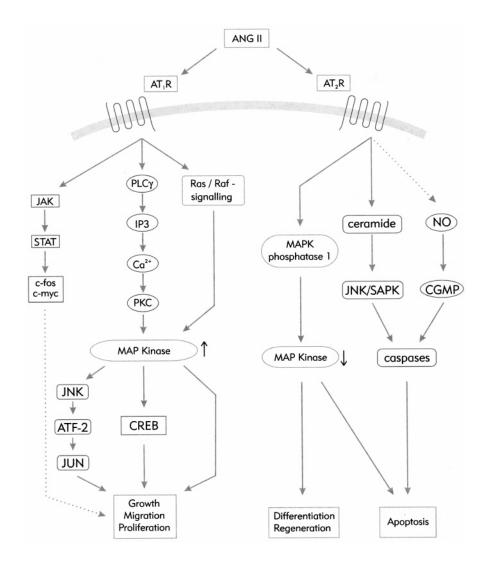


Figure 2. Angiotensin signalling pathways. Stimulation of the AT_1 receptor produces cell growth, proliferation and migration through stimulation of various kinases. AT_2 receptor stimulation induces differentiation and regeneration, although it may also cause apoptosis via ceramide or inhibition of MAPK. PLC, phospholipase C; IP3, inositol triphosphate, PKC, protein kinase C; JAK, Janus kinase; STAT, signal transducers and activators of transcription; MAPK, mitogen-activated protein kinase; JNK, c-Jun terminal kinase; for further details and abbreviations see text.

of inducible transcription factors (ITFs) such as c-Jun, c-Fos or KROX [83]. These DNA-binding proteins regulate gene expression and thus transform the signal into cellular function. ANG II is therefore involved—via the AT₁ receptor—in processes of growth, modulation and proliferation in various cell types (fig. 2). Compared to the cellular effects of AT₁ receptors, description of the AT₂-receptor-mediated signaling cascades still seems to be far from complete. After the recognition of the AT₂ receptor as a member of the G-protein-linked 7-transmembrane receptor family, several attempts failed to demonstrate G-protein-related pathways. As mentioned above, AT₂ receptor stimulation mediates antagonizing actions in terms of growth modulation. In an AT₂ receptor knock-out model, ex-

pression of AT₂ receptors was recently shown to influence fetal vascular growth via modulation of the extracellular signal-regulated kinase (ERK) activity [89]. In nervous tissue, stimulation of AT₂ receptors in cultured neurons stimulates PLA₂ activity [90], and numerous studies including tumour cell lines and neurons indicated an inhibition of MAP kinase activity and subsequent apoptosis after AT₂ receptor stimulation (fig. 2) [91, 92], pointing to involvement of an inhibitory G protein (G₁)-related mechanism. The issue is complicated by increasing evidence of cross-talk between AT₁ and AT₂ receptor signaling, e.g. the recently demonstrated AT₂-receptor-mediated inhibition of ERK and STAT activation, which is seen as a result of AT₁ receptor activation [93]. Thus, the experimental data

obtained from different cell types have further elucidated our knowledge of AT₂ receptor signaling, but some aspects are still an enigma.

AT₂ receptors and neuronal differentiation

Neuronal cell proliferation, differentiation and apoptosis are controlled by various neurotrophic factors. For example, PC12W cells, which represent an established model system for studying various aspects of neuronal differentiation and apoptosis, undergo differentiation to sympathetic neuron-like cells in response to nerve growth factor (NGF) application [94]. After growth factor withdrawal, an apoptotic programme is activated [95]. Since PC12W cells express abundant AT₂ and only few, if any, AT₁ receptors, these cells offer an excellent model to investigate AT₂-receptor-mediated effects on neuronal cells.

AT₂ receptors are not only involved in the initiation of cell differentiation by shifting cells into cell cycling arrest but, furthermore, mediate neurite extension in PC12W cells, as recently demonstrated [52]. ANG II treatment induced neurite outgrowth and enhanced the NGF-mediated morphological changes. These effects were completely suppressed by coincubation with the specific AT₂ receptor antagonist, PD 123177. Such AT₂receptor-mediated neurite extension has also been observed by Laflamme et al. [96] using NG108-15 cells, a neuroblastoma/glioma cell line. Neurite outgrowth after angiotensin treatment was also detectable in neurons derived from postnatal rat dorsal root ganglia (R. Lucius, R. Rosenstiel, S. Gallinat, J. Sievers, T. Unger unpublished data) and in postnatal retinal explants [51]. Therefore, AT₂ receptors are not only prerequisite for differentiation by exerting antiproliferative effects but appear to actively contribute to neuronal differentiation, visible e.g. by induction of neurite extension.

Neuronal differentiation is accompanied by dramatic changes in gene and protein expression. The stability and function of neurons is dependent on protein filaments like microtubules, actin filaments and intermediate filaments. These changes are reflected, for instance, in PC12W cells upon NGF treatment.

One intermediate filament protein class we were interested in were neurofilament (NF) triplet proteins, which are exclusively expressed in neurons and which can be detected at early stages of embryogenesis [97, 98]. Three subunits of NFs with molecular masses of 68 kDa (NF-L), 160 kDa (NF-M) and 200 kDa (NF-H) are known. Besides being responsible for the stability of axons, this type of intermediate filament protein is involved in axial growth of neurons following the elongation phase [99, 100], leading to increased conduction velocity [100].

To further characterize the AT_2 -receptor-induced morphological changes in PC12W cells, we investigated the effects of AT_2 receptors on the expression pattern of NF-M. In contrast to NGF stimulation [101, 102], the morphological changes after AT_2 receptor activation were paralleled by an AT_2 -receptor-mediated down-regulation of NF-M [53]. We also demonstrated that these receptors modulate the expression of MAP1B, MAP2 and β tubulin in PC12W cells [103]. MAP1B expression was attenuated, whereas AT_2 receptor stimulation upregulated β tubulin and MAP2.

These results are in line with recent results from other scientists. For example, Laflamme et al. [96] have shown that the AT_2 -receptor-induced neurite outgrowth in NG108-15 cells was accompanied by the regulation of β tubulin, tau and MAP2C, suggesting a role for AT_2 receptors in the regulation of protein filaments in neuronal cells.

The interaction of AT_2 receptors with T-type calcium channels observed in non-differentiated NG108-15 cells [81] together with the observation of the crucial role of calcium in neuronal differentiation [82] also indicate the involvement of AT_2 receptors in this process.

In summary, these data show that in different cell lines of neuronal origin, AT_2 receptor stimulation exerts antiproliferative actions and induces neurite extension. In addition to these visible morphological changes, modulation of the expression of important protein filaments through AT_2 receptor activation shows that these receptors are directly involved in neuronal development by cytoskeletal reorganization.

AT2 receptors in tissue repair and regeneration

As already mentioned, AT_2 receptors have been implicated in the process of wound healing and tissue repair [48, 49, 71]. Increased tissue AT_2 levels as early as 24 h after myocardial infarction [49] and after brain injury [54, 104] have been reported. In myocardial infarction, this effect was neither influenced by pretreatment with the AT_1 receptor antagonist, losartan, nor the ACE inhibitor, ramipril [105].

In coronary endothelial cells, both angiotensin receptor subtypes regulate several extracellular matrix proteins, i.e. fibronectin, thrombospondin and tenascin [106]. This observation not only points to the potency of both receptor subtypes to alter the adhesion and migration of coronary endothelial cells, it further suggests that angiotensin receptors are capable of promoting neurite extension by influencing the extracellular matrix composition, of which molecules like fibronectin or laminin are known to promote neurite outgrowth. Another aspect of interest in tissue repair processes is the involvement of angiotensin receptors in angiogenic processes

[107]. The authors could show that ANG II not only facilitated the activation of pre-existing collateral vessels in the rabbit cornea but, furthermore, exerted angiogenic properties. Unfortunately, the ANG-II-induced angiogenesis was not attributed to any angiotensin receptor subtype, but these effects of ANG II can be explained by AT_1 receptor stimulation, since all known growth-promoting effects of ANG II are mediated by AT_1 receptors.

However, AT₂ receptors have not only been implicated in multicellular repair systems like the process of wound healing but also in nerve regeneration after injury. Whereas the immature mammalian central nervous system (CNS) can regenerate [108], adult neurons are in most cases unable to reinnervate their target regions after injury, probably due to the actions of inhibitory molecules from CNS myelin [109] or the absence of requisite neurotrophic molecules. However, it has been shown that CNS neurons are able to regenerate new processes over long distances and to reinnervate their target region [110, 111] if they are supplied with growthpromoting substrates, e.g. transplants of fetal CNS tissue [112] or peripheral nerves [113]. The first evidence for an involvement of AT2 receptors in either apoptotic or neuroregenerative events came from our observation of the AT₂-receptor-mediated down-regulation of NF-M in PC12W cells [53]. Diminished NF-M expression has also been observed in neurons following nerve transection [114–119] and in neurons undergoing apoptosis [120]. Additionally, the modulation of MAP1B expression involves AT₂ receptors in PC12W cells [103], a process which is of interest, since down-regulation of MAP1B has been observed in regenerating neurons following axotomy [121, 122]. In view of this fact, it is intriguing to speculate about a physiological role for AT₂ receptors in these events.

Book et al. [123] detected a correlation between a decrease in MAP2 levels and death of neurons, using peripheral and central nerve lesions. Therefore, the AT_2 -receptor-induced MAP2 up-regulation in PC12W cells would support a role for AT_2 receptors in the inhibition of PCD and the promotion of regenerative processes (fig. 3).

Using a peripheral nervous system lesion approach, we could demonstrate a several-fold up-regulation of AT_2 and AT_1 receptor mRNA after sciatic nerve transection in dorsal root ganglion neurons and in sciatic nerve segments of adult rats [124]. Successful regeneration in this model under regeneration-permissive conditions, i.e. sciatic nerve crush, additionally revealed a transient up-regulation of AT_2 receptor mRNA with its expression declining in parallel with axonal elongation. These results suggest that in the peripheral nervous system, AT_2 receptors exert their actions by modulating Schwann-cell-mediated activity, e.g. the production of

neurotrophic factors, myelin degeneration or preparation of Schwann cells for subsequent synthesis of the myelin sheath.

These observations in conjunction with the above-mentioned evidence for AT₂-receptor-mediated neuronal differentiation prompted us to investigate the potential role of ANG II, acting through the AT₂ receptor, as a neurotrophic factor for CNS neurons in vitro and in vivo.

We investigated the effect of ANG II receptor stimulation on axonal regeneration of postnatal rat retinal explants and cultured dorsal root ganglia (DRG) cells in vitro and after optic nerve crush in vivo [51]. In the in vitro model, the retinal ganglion cells (RGCs) are comparable to adult, non-regenerating RGCs [125, 126]. ANG II $(10^{-7}-10^{-5} \text{ M})$ induced regeneration of neurites in a concentration-dependent manner, as shown by significant neurite outgrowth in vitro (retinal explants and the DRG cells) and in vivo after lesion of the optic nerve. The effects of ANG II—both in vitro and in vivo—were mediated by the AT₂ receptor, since (i) the effects were mimicked by CGP 42112 (an AT₂ receptor agonist), (ii) they were not suppressed by losartan (an AT₁ receptor antagonist) and (iii) they were abolished by coincubation with PD 123177 (an AT₂ receptor antagonist). The involvement of AT₂ receptors in these events was also demonstrated by a reverse transcription-polymerase chain reaction assay in which a time-dependent increase of AT₂ receptor mRNA—as in the previous studies in the peripheral nerve—could be seen both in the retina and the crushed optic nerve. These results clearly demonstrate that ANG II via its

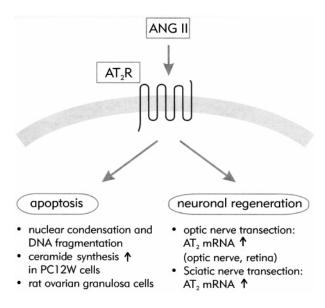


Figure 3. AT_2 receptors: apoptosis versus regeneration. For details see text.

AT₂ receptor induces axonal regeneration not only in postnatal retinal explants or DRG neurons in vitro, but also in the adult optic nerve after lesion.

Evidence for AT₂-receptor-mediated programmed cell death in neurons

Axon interruption elicits a complex neuronal response that leaves neurons poised between death and regeneration. The risk of apoptosis and the potency for axonal regeneration are closely related: mammalian CNS neurons lesioned close to their cell bodies show a strong cell body response and high regenerative capacity but, simultaneously, there is a high risk for cell death. If CNS neurons are lesioned more distally, regenerative potency is weak, but the neurons are somehow protected from apoptosis. The transcription factor c-Jun is, for example, one of the earliest markers for neurons responding to nerve fibre transection and its expression can be related to both degeneration and survival [127]. These observations suggest that neuronal injury initiates—at least in the early events of injury—a series of molecular events that are identical for both regeneration and apoptosis. Therefore, the axotomy-induced AT₂ receptor up-regulation and the AT₂ receptor-mediated NF-M down-regulation in PC12W cells [53] might also be interpreted in terms of apoptosis instead of neuroregeneration. In addition, AT₁ and AT₂ receptors were reported to be involved in modulation of the proteolytic activity of the extracellular microenvironment of neurons, e.g. by down-regulation of the serine protease inhibitor protease nexin-1 (PN-1) in primary cultures of Schwann cells [128]. PN-1 is induced after lesion of the sciatic nerve, has been demonstrated to rescue motoneurons from axotomy-induced cell death [129] and is markedly decreased in Alzheimer's disease brain [130, 131]. In PC12W cells, AT₂ receptors induce chromatin condensation and internucleosomal DNA fragmentation—molecular changes typical for apoptosis. Furthermore, the authors observed antagonization of NGF-induced MAP kinase activity by AT2 receptors [91]. Since selective inhibition of the MAP kinase pathway induces apoptosis and thus prevents survival of PC12W cells [132], AT₂ receptors were concluded to play a role in programmed cell death (PCD). Outside the nervous system, AT2 receptors have been associated with PCD in rat ovarian granulosa cells [133] and in GN4 liver epithelial cells, where ANG II was able to activate the cell-death-signaling JNK pathway [134]. To further elucidate AT₂-receptor-mediated neuronal regeneration or apoptosis, we could recently demonstrate that stimulation of the AT₂ receptor selectively induces de novo synthesis of ceramides in PC12W cells [57]. Ceramide acts as a lipid second messenger and is an important mediator of PCD. This apoptotic effect is accompanied by activation of mitogen-activated protein kinase phosphatase-1 (MKP-1) and the subsequent dephosporylation of bcl-2, an antiapoptotic protein [91], and activation of the cell death protease caspase-3 (CPP32), leading to induction of apoptosis [135]. Our finding of AT₂-receptor-mediated ceramide generation in PC12W cells—confirmed in the meantime by Lehtonen et al. [136]—connects this receptor to important apoptotic pathways.

Concluding remarks and perspectives

In this review article, we have focused on the role of the octapeptide ANG II both in regeneration/tissue repair and in apoptosis, mainly mediated via the AT₂ receptor. It appears now that the well-known growth-promoting effects of the AT₁ receptor, which can engender neuroplastic as well as pathological structural changes in several organs, are counteracted within the RAS itself by growth arrest, differentiation and tissue repair, effected through the AT₂ receptor, and a (disturbed) balance between the opposing actions of these two receptors determines the net effects of the RAS in a given disease situation. The AT₂ receptor appears to act as a modulator of complex biological programmes in development, cell differentiation, tissue repair and PCD. Many observations cited in this review could be interpreted in terms of both PCD and survival—especially neuronal survival—but at this point in time it is difficult to assess whether ANG II mediates PCD, regeneration or both.

In the CNS, the signal transduction cascades leading to either apoptosis or nerve fibre regeneration are not completely understood. How a common initial signaling pathway leads to the apparently opposed responses of cell death and axonal regeneration is a matter of intense debate

That AT₂ receptors are involved in cell differentiation processes and exert antiproliferative actions is widely accepted. However, direct in vivo evidence implicating these receptors in both neuroregeneration and PCD is still lacking. Even if participation in these events has been suggested by several studies, further investigations will have to determine precisely the physiological role of this receptor subtype. The findings on the neurotrophic actions of the AT₂ receptor may provide a basis for the design of new, receptor-directed, therapeutic strategies in the failure of axonal regeneration in the mammalian CNS. This is of particular interest considering the current difficulties in applying neurotrophic factors after nerve fibre damage. Moreover, AT₂-mediated tissue regeneration may not be confined to axonal regrowth but may constitute a general phenomenon to be exploited

by therapeutic intervention. It will be interesting to see to what extent the results obtained from neuronal cells can be applied to the cardiovascular system, under physiological and pathophysiological situations. The clinical relevance of this approach is already apparent with the increasing use of AT₁ receptor antagonists as antihypertensive drugs in, to date, more than two million patients worldwide. Since AT2 receptors are unmasked and ANG II levels increased by AT₁ receptor antagonists, part of the organ-protective actions of these drugs might be ascribed to an agonistic action of ANG II at the AT₂ receptor site. Beyond the 'classical' actions of ANG II—a story that began 100 years ago-future investigations will shed new light on the complex function of the RAS both in the CNS and periphery.

- 1 Tigerstedt R. and Bergman P. G. (1898) Niere und Kreislauf. Skand. Arch. Physiol. 8: 223-231
- 2 Unger T., Chung O., Csikos T., Culman J., Gallinat S., Gohlke P. et al. (1996) Angiotensin receptors. J. Hypertens. 14: 95–103
- 3 Timmermans P. B. M. W. M., Wong P. C., Chiu A. T., Herblin W. F., Benfield P., Carin D. J. et al. (1993) Angiotensin II receptors and angiotensin II receptor antagonists. Pharmacol. Rev. 45: 205–251
- 4 Aceto J. F. and Baker K. M. (1990) [Sar1] angiotensin II receptor-mediated stimulation of protein synthesis in chick heart cells. Am. J. Physiol. 258: H806–H813
- 5 Geisterfer A. A. T., Peach M. J. and Owens G. K. (1990) Angiotensin II induces hypertrophy but not hyperplasia of cultured rat aortic smooth muscle cells. Circ. Res. 62: 749– 756
- 6 Paquet J. C., Bandomin-Legros M., Brunelle G. and Meyer P. (1990) Angiotensin II-induced proliferation of aortic myocytes in spontaneously hypertensive rats. J. Hypertens. 8: 565-572
- 7 Stoll M., Steckelings U. M., Paul M., Bottari S. P., Metzger R. and Unger T. (1995) The angiotensin AT2 receptor mediates inhibition of cell proliferation in coronary endothelial cells. J. Clin. Invest. 95: 651–657
- 8 Owens G. K., Rabinowitch P. S. and Schwartz S. M. (1991) Smooth muscle cell hypertrophy versus hyperplasia in hypertension. Proc. Natl. Acad. Sci. USA 78: 7759–7763
- 9 Bumpus F. M., Catt K. J. and Chiu A. T. (1991) Nomenclature for angiotensin receptors: a report of the nomenclature committee of the Council for High Blood Pressure Research. Hypertension 17: 720–722
- Brechler V., Jones P. W., Levens N. R., deGasparo M. and Bottari S. P. (1993) Agonistic and antagonistic properties of angiotensin analogs at the AT2 receptor in PC12W cells. Regul. Pept. 44: 207–213
- 11 Elton T. S., Stephan C. C., Taylor G. R., Kimball M. G., Martin M. M., Durand J. N. et al. (1992) Isolation of two distinct type I angiotensin II receptor genes. Biochem. Biophys. Res. Commun. 184: 1067–1073
- 12 Ye M. Q. and Healy D. P. (1992) Characterization of an angiotensin type-1 receptor partial cDNA from rat kidney: evidence for a novel AT1b receptor subtype. Biochem. Biophys. Res. Commun. 185: 204–210
- 13 Yoshida H., Kakuchi J. and Guo D. F. (1992) Analysis of the evolution of angiotensin II type 1 receptor gene in mammals (mouse, rat, bovine, human). Biochem. Biophys. Res. Commun. **186:** 1042–1049
- 14 Ernsberger P., Zhou J., Damon T. H. and Douglas J. G. (1992) Angiotensin II receptor subtypes in cultured rat renal mesangial cells. Am. J. Physiol. 263: F411-F416

15 Siemens I. R, Reagan L. P., Yee D. K. and Fluharty S. J. (1994) Biochemical characterization of two distinct angiotensin AT2 receptor populations in murine neuroblastoma N1E-115 cells. J. Neurochem. 62: 2106-2115

- 16 Swanson G. N., Hannesworth J. N. and Sardinia M. F. (1992) Discovery of a distinct binding site for angiotensin (3-8) a putative angiotensin IV receptor. Regul. Pept. 40: 409-419
- 17 Harding J. W., Wright J. W., Swanson G. N., Hanesworth J. M. and Krebs L. T. (1994) AT4 receptors: specificity and distribution. Kidney Int. 46: 1510–1512
- 18 Wang L., Eberhard M. and Erne P. (1995) Stimulation of cDNA and RNA synthesis of cultured rabbit cardiac fibroblasts by angiotensin IV. Eur. Heart J. 16 [suppl]: 451
- 19 Roberrs K. A., Krebs L. T., Kramar E. A., Shaffer M. J., Harding J. W. and Wright J. W. (1995) Autoradiographic identification of brain angiotensin IV binding sites and differential c-fos expression following intracerebroventricular injection of angiotensin II and IV in rats. Brain Res. 682: 13-21
- 20 Neuß M., Regitz-Zagrosek V. and Fleck E. (1994) Human cardiac fibroblasts express an angiotensin receptor with unusual binding characteristics. Biochem. Biophys. Res. Commun. 204: 1334–1339
- 21 Eggena P., Zhu J. H., Clegg K. and Barrett J. D. (1993) Nuclear angiotensin receptors transduce transcription of renin and angiotensin mRNA. Hypertension 22: 496–501
- 22 Chiu A. T., Herlin W. F. and McCall D. E. (1989) Identification of angiotensin II receptor subtypes. Biochem. Biophys. Res. Commun. 165: 196–203
- 23 Gehlert D. R., Gachenheimer S. L. and Schober D. A. (1991) Autoradiographic localization of subtypes of angiotensin II antagonist binding in the rat brain. Neuroscience 44: 501–514
- 24 Millan M. A., Jacobowitz D. M., Aguilera G. and Catt K. J. (1991) Differential distribution of AT1 and AT2 angiotensin II receptor subtypes in the rat brain during development. Proc. Natl. Acad. Sci. USA 88: 11440–11444
- Obermüller N., Unger T., Culman J., Gohlke P., deGasparo M. and Bottari S. P. (1991) Distribution of angiotensin II receptor subtypes in rat brain nuclei. Neurosci. Lett. 132: 11-15
- 26 Saveedra J. M (1992) Brain and pituitary angiotensin. Endocrinol. Rev. 13: 329–380
- 27 Steckelings U., Obermüller N., Bottari S. P., Quadri F., Veltmar A. and Unger T. (1992) Brain angiotensin: receptors, actions and possible role in hypertension. Pharmacol. Toxicol. 70 [suppl. II]: 23–27
- 28 Tsutsumi K. and Saveedra J. M. (1991) Characterization and development of angiotensin II receptor subtypes (AT₁ and AT₂) in rat brain. Am. J. Physiol. 216: R209–R216
- 29 Whitebread S., Mele M., Kamber B. and deGasparo M. (1989) Preliminary biochemical characterization of two angiotensin II receptor subtypes. Biochem. Biophys. Res. Commun. 163: 284–291
- 30 Zhuo J., Song K., Harris P. J. and Mendelsohn F. A. O. (1992) In vitro autoradiography reveals predominantly AT1 angiotensin II receptors in rat kidney. Renal Physiol. Biochem. 15: 231–239
- 31 Bauer P. H., Chiu A. T. and Garrison J. C. (1991) DuP 753 can antagonize the effects of angiotensin II in rat liver. Mol. Pharmacol. **39:** 579–585
- 32 Chang R. S. L. and Lotti V. J. (1989) Selective ligands reveal subtypes of angiotensin receptors in rat vasculature and brain. Pharmacologist 31: 150
- 33 Chang R. S. L. and Lotti V. J. (1990) Two distinct angiotensin II receptor binding sites in rat adrenal revealed by new selective nonpeptide ligands. Mol. Pharmacol. 29: 347–351
- 34 Chang R. S. L. and Lotti V. J. (1991) Angiotensin receptor subtypes in rat, rabbit and monkey tissues: relative distribution and species dependency. Life Sci. 49: 1485–1490

- 35 Chappell M. C., Diz D. I and Jacobson D. W. (1992) Pharmacological characterization of angiotensin II binding sites in the canine pancreas. Peptides 13: 313–318
- 36 Dudley D. T., Panek R. L. and Major T. C. (1990) Subclasses of angiotensin II binding sites and their functional significance. Mol. Pharmacol. 38: 370–377
- 37 Dudley D. T., Hubbel S. E. and Summerfelt R. M. (1991) Characterization of angiotensin II (AT2) binding sites in R3T3 cells. Mol. Pharmacol. 40: 360–367
- 38 Eglen R. M., Rapp J. M. and Leung E. (1991) Characterization of angiotensin II receptors in guinea pig gastrointestinal tract. FASEB J. 5: A869
- 39 Entzeroth M. and Hadamovsky S. (1991) Angiotensin II receptors in the rat lung are of the AII-1 subtype. Eur. J. Pharmacol. 206: 237-241
- 40 deGasparo M., Whitebread S. and Mele M. (1990) Biochemical characterization of two angiotensin II receptor subtypes in the rat. J. Cardiovasc. Pharmacol. 16 [suppl. 4]: S31–S35
- 41 Gibson R. E, Thorpe H. H. and Cartwright M. E. (1991) Angiotensin II receptor subtypes in renal cortex of rats and rhesus monkeys. Am. J. Physiol. 261: F512–F518
- 42 Herblin W. F., Chiu A. T. and McCall D. E. (1991) Angiotensin II receptor heterogeinity. Am. J. Hypertens. 4: 299S-302S
- 43 Pucell A. G., Hodges J. C., Sen I., Bumpus F. M. and Husain A. (1991) Biochemical properties of the ovarian granulosa cell type 2 angiotensin II receptor. Endocrinology 128: 1947–1959
- 44 Schinke M., Doods H.N., Ganten D., Wienen W. and Entzeroth M. (1991) Characterization of rat intestinal angiotensin II receptors. Eur. J. Pharmacol. 204: 165–170
- 45 Simon M., Flügge A., Fuchs E. and Gröne H. J. (1991) Evidence for two angiotensin II receptor subtypes in human fetal and adult kidney demonstrated by the AII receptor antagonist losartan and PD123177. FASEB J. 5: A870
- 46 Tsutsumi K., Strömberg C., Viswanathan M. and Saveedra J.M. (1991) Angiotensin II receptor subtypes in fetal tissue of the rat: autoradiography, guanine nucleotide sensitivity, and association with phosphoinoside hydrolysis. Endocrinology 129: 1075–1082
- 47 Grady E. F., Sechi L. A., Griffin C. A., Schambelan M. and Kalinyak J. E. (1991) Expression of AT2 receptors in the developing rat fetus. J. Clin. Invest. 88: 921–933
- 48 Kimura B., Sumners C. and Phillips M. I. (1992) Changes in skin angiotensin II receptors in rats during wound healing. Biochem. Biophys. Res. Commun. 187: 1083–1090
- 49 Nio Y., Matsubara H., Murasawa S., Kanasaki M. and Inada M. (1995) Regulation of gene transcription of angiotensin II receptor subtypes in myocardial infarction. J. Clin. Invest. 95: 46–54
- 50 Gallinat S., Yu M. H., Dorst A., Unger T. and Herdegen T. (1998) Sciatic nerve transection evokes lasting up-regulation of angiotensin AT2 and AT1 receptor mRNA in adult rat dorsal root ganglia and sciatic nerves. Mol. Brain Res. 57: 111-122
- 51 Lucius R., Gallinat S., Rosenstiel P., Herdegen T., Sievers J. and Unger T. (1998) The angiotensin II type 2 (AT2) receptor promotes axonal regeneration in the optic nerve of adult rats. J. Exp. Med. 188: 661–670
- 52 Meffert S., Stoll M., Steckelings U. M., Bottari S. P. and Unger T. (1996) The angiotensin AT2 receptor inhibits proliferation and promotes differentiation in PC12W cells. Mol. Cell. Endocrinol. 122: 59–67
- 53 Gallinat S., Csikos T., Meffert S., Herdegen T., Stoll M. and Unger T. (1997) The angiotensin AT2 receptor down-regulates neurofilament M in PC12W cells. Neurosci. Lett. 227: 29-32
- 54 Viswanathan M. and Saavedra J. M. (1992) Angiotensin II AT2 receptor in the rat brain during experimental wound healing. Peptides 13: 783-786
- Makino I., Shibata K., Shibaguchi H., Niwa M., Katsuragi T. and Furukawa T. (1998) The increase in angiotensin type-2 receptor mRNA level by glutamate stimulation in cultured rat cortical cells. Brain Res. 804: 296-305

- 56 Shibata K., Makino I., Shibaguchi H., Niwa M., Ohgami Y., Fujiwara M. et al. (1998) Expression of angiotensin type-2 receptors in rat brain during cell injury. Nippon Yakurigaku Zasshi 112 [suppl 1]: 53P-57P
- 57 Gallinat S., Busche S., Schütze S., Krönke M. and Unger T. (1999) AT2 receptor stimulation induces generation of ceramides in PC12W cells. FEBS Lett. 443: 75–79
- 58 Shenoy U. V., Richards E. M., Huang X. C. and Sumners C. (1999) Angiotensin type 2 receptor-mediated apoptosis of cultured neurons from newborn rat brain. Endocrinology 140: 500-509
- Phillips M. I., Weyhenmeyer J. A., Felix D. and Ganten D. (1979) Evidence of an endogenous brain renin angiotensin system, Fed. Proc. 38: 2260–2266
- 60 Ganten D., Hermann K., Bayer C., Unger T. and Lang R. E. (1983) Angiotensin synthesis in the brain and increased turnover in hypertensive rats. Science 221: 869–871
- 61 Lynch K. R., Simnad V. I., Ben-Ari E. T. and Garrison J. C. (1986) Localization of angiotensinogen messenger RNA sequences in the rat brain. Hypertension 8: 540–543
- 62 Dzau V. J., Ingelfinger J., Pratt R. E. and Ellison K. E. (1986) Identification of renin and angiotensinogen messenger RNA sequences in the rat brain. Hypertension 8: 544–548
- 63 Song K., Allen A. M., Paxinos G. and Mendelsohn F. A. O. (1992) Mapping of angiotensin II receptor subtype heterogeneity in rat brain. J. Comp. Neurol. 316: 467–484
- 64 Goldberg A. (1995) Safety and tolerability of losartan potassium, an angiotensin II receptor antagonist, compared with hydrochlorothiazid, atenolol, felodipine ER, and angiotensin-converting enzyme inhibitors for the treatment of hypertension. Am. J. Cardiol. 75: 793–795
- 65 Camargo M. J. F., Lutterotti N. von, Campell W. G. Jr, Pecker M. S., James G. D., Timmermanns P. B. M. W. M. et al. (1993) Control of blood-pressure and end-organ damage in maturing salt-loaded stroke prone spontaneously hypertensive rats by oral angiotensin II receptor blockade. J. Hyperten. 11: 31–40
- 66 Fornes P., Richer C., Vacher E., Bruneval P. and Giudicelli J. F. (1993) Losartan's protective effects in stroke prone spontaneously hypertensive rats persist durably after treatment withdrawal. J. Cardiovasc. Pharmacol. 22: 305–313
- 67 Stie C. T. Jr, Adler L. A., Levine S. and Chander P. N. (1993) Stroke prevention by losartan in stroke prone spontaneously hypertensive rats. J. Hyperten. 11 [suppl]: 37–42
- 68 Gohlke P., Linz W., Schölkens B. A., Wiemer G. and Unger T. (1996) Cardiac and vascular effects of long-term losartan treatment in stroke prone spontaneously hypertensive rats. Hypertension 3: 397–402
- 69 Sachinidis A., el-Haschimi K., Ko Y., Seul C., Dusing R. and Vetter H. (1996) CV-11974, the active metabolite of TCV-116 (candesartan), inhibits the synergistic or additive effect of different growth factors of angiotensin II-induced proliferation of vascular smooth muscle cells. Biochem. Pharmacol. **52**: 123–126
- 70 Yanagitani Y., Rakugi H., Okamura A., Moriguchi K., Takiuchi S., Ohishi M. et al. (1999) Angiotensin II type 1 receptor-mediated peroxide production in human macrophages. Hypertension 33: 335–339
- 71 Nakajima M., Hutchinson H. G., Fujinaga M., Hayashida W., Morishita R., Zhang L. et al. (1995) The angiotensin II type 2 (AT2) receptor antagonizes the growth effects of the AT1 receptor: gain-of-function study using gene transfer. Proc. Natl. Acad. Sci. USA 92: 10663-10667
- 72 Munzenmaier D. H. and Greene A. S. (1996) Opposing actions of angiotensin II on microvascular growth and arterial blood pressure. Hypertension 27: 760–765
- 73 Hein L., Barsh G. S., Pratt R. E., Dzau V. J. and Kobilka B. K. (1995) Behavioural and cardiovascular effects of disrupting the angiotensin II type-2 receptor gene in mice. Nature 377: 744–747
- 74 Ichiki T., Labosky P. A., Shiota C., Okuyama S., Imagawa Y., Fogo A. et al. (1995) Effects of blood pressure and exploratory behaviour of mice lacking angiotensin II type-2 receptor. Nature 377: 748–750

75 Kitamura Y., Sasamura H., Maruyama T., Nakaya H., Amemiya T., Hayashi M. et al. (1998) Adrenal angiotensin II type 1 and type 2 receptors in Cushing's and Conn's syndromes. Mol. Cell. Endocrinol. 144: 37–45

- 76 Tsutsumi K., Viswanathan M., Stromberg C. and Saavedra J. M. (1991) Type-1 and type-2 angiotensin II receptors in fetal rat brain. Eur. J. Pharmacol. 198: 89–92
- 77 Leung K. H., Roscoe W. A., Smith R. D., Timmermans P. B. M. W. M and Chiu A. T. (1991) Regional distribution of two subtypes of angiotensin II receptor in rat brain using selective nonpeptide antagonists. Neurosci. Lett. 123: 95–98
- 78 Speth R. C., Mei L. and Yamamura H. I. (1989) Angiotensin II receptor binding and actions in NG 108-15 cells. Peptide Res. 2: 232-239
- 79 Webb M. L., Liu E. C. K., Cohen R. B., Hedberg A., Bogosian E. A., Monshizadegan C. M. et al. (1992) Molecular characterization of angiotensin type II receptors in rat pheochromocytoma cells. Peptides 13: 499–508
- 80 Buisson B., Bottari S. P., Gasparo M. de, Gallo-Payet N. and Payet M. D. (1992) The angiotensin AT2 receptor modulates T-type calcium current in non-differentiated NG 108-15 cells. FEBS Lett. 309: 161-164
- 81 Buisson B., Laflamme L., Bottari S. P., Gasparo M. de, Gallo-Payet N. and Payet M. D. (1995) A G-protein is involved in the angiotensin AT2 receptor inhibition of the T-type Ca²⁺ current in non-differentiated NG 108-15 cells. J. Biol. Chem. 270: 1670–1674
- 82 Kater S. B. and Mills L. R. (1991) Regulation of growth cone behaviour by calcium. Neuroscience 11: 891–899
- 83 Blume A. and Unger T. (1999) Angiotensin peptides and inducible transcription factors. J. Mol. Med. 77: 339–357
- 84 Garcia-Sainz J. A. and Macias-Silvas M. (1990) Angiotensin II stimulates phosphoinositide turnover and phosphorylase through AII-1 receptors in isolated hepatocytes. Biochem. Biophys. Res. Commun. 172: 780–784
- 85 Balla T., Baukal A. J., Eng S. and Catt K. J. (1991) Angiotensin II receptor subtypes and biological responses in the adrenal cortex and medulla. Mol. Pharmacol. 40: 401– 406
- 86 Marrero M. B., Paxton W. B., Schieffer B., Ling B. N. and Bernstein K. E. (1996) Angiotensin II signaling events mediated by tyrosine phosphorylation. Cell Signalling 8: 21–26
- 87 Gelband C. H., Sumners C., Lu D. and Raizada M. K. (1998) Angiotensin receptors and norepinephrine neuromodulation: implications of functional coupling. Regul. Peptides 73: 141–147
- 88 Lu D., Yang H., Lenox R. H. and Raizada M. K. (1998) Regulation of angiotensin II-induced neuromodulation by MARCKS in brain neurons. J. Cell Biol. 142: 217–227
- 89 Akishita M., Itoh M., Lehtonen J. Y., Daviet L., Dzau V. J. and Horiuchi M. (1999) Expression of the AT2 receptor developmentally programs extracellular signal-regulated kinase activity and influences fetal vascular growth. J. Clin. Invest. 103: 63–71
- 90 Zhu M., Gelband C. H., Moore J. M., Posner P. and Sumners C. (1998) Angiotensin II type 2 receptor stimulation of neuronal delayed rectifier potassium currents involves phospholipase A₂ and arachidonic acid. J. Neurosci. 18: 679-686
- 91 Yamada T., Horiuchi M. and Dzau V.J. (1996) Angiotensin II type 2 receptor mediates programmed cell death. Proc. Natl. Acad. Sci. USA 93: 156–160
- 92 Horiuchi M., Hayashida W., Kambe T., Yamada T. and Dzau V. J. (1997) Angiotensin type 2 receptor dephosphorylates Bcl-2 by activating mitogen-activated protein kinase phosphatase-1 and induces apoptosis. J. Biol. Chem. 272: 19022–19026
- 93 Horiuchi M., Hayashida W., Akishita M., Tamura K., Daviet L., Lehtonen J.Y. et al. (1999) Stimulation of different subtypes of angiotensin II receptors, AT1 and AT2 receptors, regulates STAT activation by negative crosstalk. Circ. Res. 84: 876–882

- 94 Greene L. A. and Tischler A. S. (1982) PC12 pheochromocytoma cultures in neurobiological research. Adv. Cell. Neurobiol. 3: 373–414
- 95 Mesner P. W., Epting C. L., Hegarty J. L. and Green S. H. (1995) A timetable of events during programmed cell death induced by trophic factor withdrawal from neuronal PC12 cells. J. Neurosci. 15: 7357–7366
- 96 Laflamme L., Gasparo M. de, Gallo J. M., Payet M. D. and Gallo-Payet N. (1996) Angiotensin II induction of neurite outgrowth by AT2 receptors in NG108-15 cells. J. Biol. Chem. 271: 22729–22735
- 97 Tapscott S. J., Bennett G. S. and Holtzer H. (1981) Neuronal precursor cells in the chick neural tube express neurofilament proteins. Nature 292: 836–838
- 98 Cochard P. and Paulin D. (1984) Initial expression of neurofilaments and vimentin in the central and peripheral nervous system of the mouse embryo in vivo. J. Neurosci. 4: 2080–2094
- 99 Hoffman P., Cleveland D., Griffin J., Landes P., Cowan N. and Price D. (1987) Neurofilament gene expression: a major determinant of axonal caliber. Proc. Natl. Acad. Sci. USA 84: 3472–3476
- 100 Sakaguchi T., Okada M., Kitamura T. and Kawasaki K. (1993) Reduced diameter and conduction velocity of myelinated fibers in the sciatic nerve of a neurofilament-deficient mutant quail. Neurosci. Lett. 153: 65–68
- 101 Lindenbaum M. H., Carbonetto S., Grosveld F., Flavell D. and Mushynski W. E. (1988) Transcriptional and post-transcriptional effects of nerve growth factor on expression of the three neurofilament subunits in PC12 cells. J. Biol. Chem. 263: 5662-5667
- 102 Lee V., Trojanowski J. Q. and Schlaepfer W. W. (1982) Induction of neurofilament triplet proteins in PC12 cells by nerve growth factor. Brain Res. 238: 169-180
- 103 Stroth U., Meffert S., Gallinat S. and Unger T. (1997) Angiotensin II induced differentiation via AT2 receptors differs from NGF-dependent mechanisms. J. Hypertension 15: S64
- 104 Viswanathan M., De Oliveira A. M., Wu R. M., Chiueh C. C. and Saveedra J. M. (1994) [125 I]CGP42112 reveals a non-angiotensin II binding site in 1-methyl-4-phenylpyridine (MPP+)-induced brain injury. Cell. Mol. Neurobiol. 14: 99-104
- 105 Zhu Y. Z., Li. J., Zhu Y. C., Chung O., Spitznagel H., Schäfer H. et al. (1996) Gene expression of angiotensin AT1 and AT2 receptors in cardiac tissue after myocardial infarction. Hypertension 28: 694
- 106 Fischer J., Stoll M. and Unger T. (1996) Thrombospondin mRNA expression is increased in endothelial cells following stimulation of AT2 receptors. J. Vasc. Res. 33 [S1]: 26
- 107 Fernandez L. A., Twickler J. and Mead A. (1985) Neovascularization produced by angiotensin II. J. Lab. Clin. Med. 105: 141–145
- 108 Shewan D., Berry M. and Cohen J. (1995) Extensive regeneration in vitro by early embryonic neurons on mature and adult CNS tissue. J. Neurosci. 15: 2057–2062
- 109 Schwab M., Kapfhammer J. P. and Bandtlow C. E. (1993) Inhibitors of neurite growth. Annu. Rev. Neurosci. 16: 565– 505
- 110 Björklund A. and Stenevi U. (eds) (1985) Neural Grafting in the Mammalian CNS. Elsevier, Amsterdam
- 111 Aguayo A. J. (1985) Axonal regeneration from injured neurons in the adult mammalian nervous system. In: Synaptic Plasticity, pp. 457–484, Cotman C. W. (ed.), Guilford, New York
- 112 Hausmann B., Sievers J., Hermanns J. and Berry M. (1989) Regeneration of axons from the adult rat optic nerve: influence of fetal brain grafts, laminin and artificial basement membrane. J. Comp. Neurol. 281: 447–466
- 113 David S. and Aguayo A. (1981) Axonal elongation into peripheral nervous system "bridges" after central nervous system injury in adult rats. Science 214: 931–933

- 114 Hoffman P. N., Pollock S. C. and Striph G. G. (1993) Altered gene expression after optic nerve transection: reduced neurofilament expression as a general response to axonal injury. Exp. Neurol. 119: 32–36
- 115 Goldstein M. E., Weiss S. R., Lazzarini R. A., Shneidman P. S., Lees J. F. and Schlaepfer W. W. (1988) mRNA levels of all three neurofilament proteins decline following nerve transection. Brain Res. 427: 287-291
- 116 Wong J. and Oblinger M. M. (1987) Changes in neurofilament gene expression occur after axotomy of dorsal root ganglion neurons: an in situ hybridization study. Metab. Brain Dis. 2: 291–303
- 117 Muma N. A., Slunt H. H. and Hoffman P. N. (1991) Postnatal increases in neurofilament gene expression correlate with the radial growth of axons. J. Neurocytol. 20: 844–854
- 118 Schwartz M. L., Shneidman P. S., Bruce J. and Schlaepfer W. W. (1992) Actinomycin prevents the destabilization of neurofilament mRNA in primary sensory neurons. J. Biol. Chem. 267: 24596–24600
- 119 Jiang Y. Q., Pickett J. and Oblinger M. M. (1994) Comparison of changes in beta-tubulin and NF gene expression in rat DRG neurons under regeneration-permissive and regeneration-prohibitive conditions. Brain Res. 637: 233–241
- 120 Estus S., Zaks W. J., Freeman R. S., Gruda M., Bravo R. and Johnson E. M. (1994) Altered gene expression in neurons during programmed cell death: identification of c-jun as necessary for neuronal apoptosis. J. Cell Biol. 127: 1717–1727
- 121 Woodhams P. L., Calvert R. and Dunnet S. B. (1989) Monoclonal antibody G10 against microtubule-associated protein 1x distinguishes between growing and regenerating axons. Neuroscience **28**: 49–59
- 122 Svensson M. and Aldskogius H. (1992) The effect of axon injury on microtubule-associated proteins MAP2, 3 and 5 in the hypoglossal nucleus of the adult rat. J. Cytol. **21:** 222–231
- 123 Book A. A., Fischer I., Yu X. J., Lannuzelli P. and Murphy E. H. (1996) Altered expression of microtubule-associated proteins in cat trochlear motoneurons after peripheral and central lesions of the trochlear nerve. Exp. Neurol. 138: 214–226
- 124 Gallinat S., Yu M. H., Csikos T., Herdegen T. and Unger T. (1997) The angiotensin AT2 receptor is involved in neuroregenerative processes. J. Hypertens. 15: S104
- 125 Allcutt D., Berry M. and Sievers J. (1984) A qualitative comparison of the reaction of retinal ganglion cells to optic

- nerve crush in neonatal and adult mice. Dev. Brain Res. 16: 231-240
- 126 Allcutt D., Berry M. and Sievers J. (1984) A quantitative comparision of the reaction of retinal ganglion cells to optic nerve crush in neonatal and adult mice. Dev. Brain Res. 16: 219–230
- 127 Herdegen T., Skene P. and Bähr M. (1997) The c-Jun transcription factor bipotential mediator of neuronal death, survival and regeneration. Trends Neurosci. 20: 227–231
- 128 Bleuel A., deGasparo M., Whitebread S., Puttner I. and Monard D. (1995) Regulation of protease nexin-1 expression in cultured Schwann cells is mediated by angiotensin II receptors. J. Neurosci. 15: 750-761
- 129 Houenou L. J., Turner P. L., Li L., Oppenheim R. W. and Festoff B. W. (1995) A serine protease inhibitor, protease nexin-1, rescues motoneurons from naturally occurring and axotomy-induced cell death. Proc. Natl. Acad. Sci. USA 92: 895–899
- 130 Choi B. H., Kim R. C., Vaughan P. J., Lau A., Van Nostrand W. E., Cotman C. W. et al. (1995) Decreases in protease nexins in Alzheimer's disease brain. Neurobiol. Aging 16: 557–562
- 131 Vaughan P. J., Su J., Cotman C. W. and Cunningham D. D. (1994) Protease nexin-1, a potent thrombin inhibitor, is reduced around cerebral blood vessels in Alzheimer's disease. Brain Res. 668: 160–170
- 132 Kummer J. L., Rao P. K. and Heidenreich K. A. (1997) Apoptosis induced by withdrawal of trophic factors is mediated by p38 mitogen-activated protein kinase. J. Biol. Chem. 272: 20490–20494
- 133 Tanaka M., Ohnishi J., Ozawa Y., Sugimoto M., Usuki S. and Naruse M. (1995) Characterization of angiotensin II receptor type 2 during differentiation and apoptosis of rat ovarian cultured granulosa cells. Biochem. Biophys. Res. Commun. 207: 593-598
- 134 Zohn I. E., Yu H., Li X., Cox A. D. and Earp H. S. (1995) Angiotensin II stimulates calcium-dependent activation of c-Jun N-terminal kinase. Mol. Cell. Biol. 15: 6160-6168
- 135 Dimmeler S., Rippmann V., Weiland U., Haendeler J. and Zeiher A. M. (1997) Angiotensin II induces apoptosis of human endothelial cells: protective effect of nitric oxide. Circ. Res. 81: 970-976
- 136 Lehtonen J. Y., Horiuchi M., Daviet L., Akishita M. and Dzau V. J. (1999) Activation of the de novo biosynthesis of sphingolipids mediates angiotensin II type 2 receptor-induced apoptosis. J. Biol. Chem. 274: 16901–16906